

A NOVEL PRESENILIN ASSOCIATED MEMBRANE PROTEIN AND USES THEREOF

This application is a divisional of U.S. Serial No. 09/541,094, filed
5 March 31, 2000, which claims the priority of U.S. provisional patent application Serial Nos.
60/127,452 filed April 1, 1999 and 60/173,826, filed December 30, 1999, each of which is
hereby incorporated herein by reference.

FIELD OF THE INVENTION

The present invention relates generally to the field of neurological and
10 physiological dysfunctions associated with neuropsychiatric and developmental diseases,
especially Alzheimer's Disease. More particularly, the invention is concerned with the
identification of proteins associated with neuropsychiatric and developmental diseases,
especially Alzheimer's Disease, and relates to methods of diagnosing these diseases, and to
methods of screening for candidate compounds which modulate the interaction of a certain
15 protein, specifically Presenilin Associated Membrane Protein ("PAMP"), with presenilin
proteins.

BACKGROUND OF THE INVENTION

Alzheimer's Disease (AD) is a degenerative disorder of the human central
20 nervous system characterized by progressive memory impairment and cognitive and intellectual
decline during mid to late adult life (Katzman, *N Eng J Med* 1986;314:964-973). The disease
is accompanied by a constellation of neuropathologic features principal amongst which are the
presence of extracellular amyloid or senile plaques, and neurofibrillary tangles in neurons.
The etiology of this disease is complex, although in some families it appears to be inherited as
25 an autosomal dominant trait. Genetic studies have identified three genes associated with the
development of AD, namely: (1) β -amyloid precursor protein (β APP) (Chartier-Harlin *et al.*,
Nature 1991;353:844-846; Goate *et al.*, *Nature* 1991;349:704-706; Murrell *et al.*, *Science*
1991;254:97-99; Karlinsky *et al.*, *Neurology* 1992;42:1445-1453; Mullan *et al.*, *Nature*
Genetics 1992;1:345-347); (2) presenilin-1 (PS1) (Sherrington *et al.*, *Nature* 1995;375:754-
30 760); and (3) presenilin-2 (PS2) (Rogaev *et al.*, *Nature* 1995;376:775-778; Levy-Lehad *et al.*,
Science 1995; 269:970-973).

Abnormal processing of β APP with overproduction of amyloid- β is also a
feature of other CNS diseases, including inherited and sporadic forms of amyloid angiopathy,

which presents with intracerebral bleeding (Levy *et al.*, *Science* 1990;248:1124-1126). Thus, abnormalities of presenilin proteins and PS-interacting proteins may affect these diseases as well.

The presenilin genes (PS1 - *PS1* and PS2 - *PS2*) encode homologous polytopic transmembrane proteins that are expressed at low levels in intracellular membranes including the nuclear envelope, the endoplasmic reticulum, the Golgi apparatus and some as yet uncharacterized intracytoplasmic vesicles in many different cell types including neuronal and non-neuronal cells (Sherrington *et al.*, *supra*; Rogaev *et al.*, *supra*; Levy-Lahad *et al.*, *supra*; Doan *et al.*, *Neuron* 1996;17:1023-1030; Walter *et al.*, *Molec. Medicine* 1996;2:673-691; De Strooper *et al.*, *J. Biol. Chem.* 1997;272:3590-3598; Lehmann *et al.*, *J. Biol. Chem.* 1997;272:12047-12051; Li *et al.*, *Cell* 1997;90:917-927). Structural studies predict that the presenilins contain between six and eight transmembrane (TM) domains organized such that the N-terminus, the C-terminus, and a large hydrophilic loop following the sixth TM domain are located in the cytoplasm or nucleoplasm, while the hydrophilic loop between TM1 and TM2 is located within the lumen of membranous intracellular organelles (Doan *et al.*, 1996; De Strooper *et al.*, 1997; Lehmann *et al.*, 1997).

Missense mutations in the PS1 and PS2 genes are associated with the inherited forms of early-onset AD (Sherrington *et al.*, *Nature* 1995;375:754-760; Rogaev, *et al.*, *Nature* 1995;376:775-778; Levy-Lahad *et al.*, *Science* 1995;269:970-973). Several lines of evidence have also suggested roles in developmental, apoptotic signalling and in the regulation of proteolytic cleavage of the β -amyloid precursor protein (β APP) (Levitan *et al.*, *Nature* 1995;377:351-354; Wong *et al.*, *Nature* 1997;387:288-292; Shen *et al.*, *Cell* 1997;89:629-639; Wolozin *et al.*, *Science* 1996;274:1710-1713; De Strooper *et al.*, *Nature* 1998;391:387-390). Nevertheless, it remains unclear just how these putative functions are mediated, or how they relate to the abnormal metabolism of the β APP associated with PS1 and PS2 mutations (Martin *et al.*, *NeuroReport* 1995;7:217-220; Scheuner *et al.*, *Nature Med.* 1996;2:864-870; Citron *et al.*, *Nature Med.* 1997;3:67-72; Duff *et al.*, *Nature* 1996;383:710-713; Borchelt *et al.*, *Neuron* 1996;17:1005-1013).

PS1 and PS2 interact specifically with at least two members of the *armadillo* family of proteins; neuronal plakophilin-related armadillo protein (Paffenholtz *et al.*, *Differentiation* 1997; 61: 293-304; Paffenholtz *et al.*, *Exp Cell Res* 1999; 250: 452-464; Zhou *et al.*, *Neuroreport* 1997; 8: 2085-2090) and β -catenin, that are expressed in both embryonic and post-natal tissues. Moreover, the domains of PS1 and PS2 that interact with these proteins have been identified. Mutations in PS1 and PS2 affect the translocation of β -catenin into the nucleus of both native cells and cells transfected with a mutant PS gene. These interactions and effects are described in detail in co-pending commonly assigned U.S. Application Serial

No. 09/227,725, filed January 8, 1999, the disclosure of which is incorporated herein by reference.

The identification and cloning of normal as well as mutant PS1 and PS2 genes and gene products are described in detail in co-pending commonly assigned US Application
5 Serial Nos. 08/431,048, filed April 28, 1995; 08/496,841, filed June 28, 1995; 08/509,359, filed July 31, 1995; and 08/592,541, filed January 26, 1996, the disclosures of which are incorporated herein by reference.

There is speculation that onset of AD may be associated with aberrant
interactions between mutant presenilin proteins and normal forms of PS-interacting proteins,
10 and these changes may increase or decrease interactions present with normal PS1, or cause interaction with a mutation-specific PS-interacting protein. Such aberrant interactions also may result from normal presenilins binding to mutant forms of the PS-interacting proteins. Therefore, mutations in the PS-interacting proteins may also be implicated in the development of AD.

15 While the identification of normal and mutant forms of PS proteins has greatly facilitated development of diagnostics and therapeutics, a need exists for new methods and reagents to more accurately and effectively diagnose and treat AD. In addition, further insights into PS proteins and their interaction with other components may lead to new diagnostic and treatment methods for other related CNS diseases.

SUMMARY OF THE INVENTION

Applicants have discovered that PS1 and PS2 interact specifically with a transmembrane protein, herein referred to as "Presenilin Associated Membrane Protein" or "PAMP", which is expressed in multiple tissues (*e.g.*, brain, kidney, lung, etc.). The product
25 of this gene is therefore implicated in the biochemical pathways affected in Alzheimer's Disease, and may also have a role in other dementias, amyloid angiopathies, and developmental disorders such as spina bifida. This gene, therefore, presents a new therapeutic target for the treatment of Alzheimer's Disease and other neurologic diseases. In addition, PAMP nucleic acids, proteins and peptides, antibodies to PAMP, cells transformed with
30 PAMP nucleic acids, and transgenic animals altered with PAMP nucleic acids that possess various utilities, as described herein for the diagnosis, therapy and continued investigation of Alzheimer's Disease and other neurodegenerative disorders. Furthermore, mutant PAMP nucleic acids, proteins, or peptides, cells transfected with vectors comprising mutant PAMP nucleic acids, transgenic animals expressing mutant PAMP or peptides thereof, and their use in
35 studying Alzheimer's Disease and other neurodegenerative disorders, or developing improved diagnostic or therapeutic methods for such disorders, are presented herein.

Thus, the invention provides isolated and purified presenilin associated membrane protein (PAMP), or a functional fragment thereof, as well as nucleic acids encoding a PAMP. Preferred nucleotide and amino acid sequences are provided herein. The invention further provides probes and primers for PAMP genes. Preferred embodiments include sequences of at least 10, 15 or 20 consecutive nucleotides selected from the disclosed sequences.

The invention also provides isolated and purified mutant PAMP, or a functional fragment thereof, as well as nucleic acids encoding a mutant PAMP, and probes and primers for PAMP genes. Preferred nucleotide and amino acid sequences are provided herein.

Using the nucleic acid and amino acid sequences disclosed herein, methods for identifying allelic variants or heterospecific homologues of a human PAMP and gene are provided. The methods may be practiced using nucleic acid hybridization or amplification techniques, immunochemical techniques, or any other technique known in the art. The allelic variants may include other normal human alleles as well as mutant alleles of PAMP genes which may be causative of Alzheimer's Disease or other CNS diseases. The heterospecific homologues may be from other mammalian species, such as mice, rats, dogs, cats or non-human primates, or may be from invertebrate species, such as *Drosophila melanogaster* or *Caenorhabditis elegans*. Thus, it is another object of the invention to provide nucleic acids that encode allelic or heterospecific variants of the disclosed sequences, as well as the allelic or heterospecific proteins encoded by them.

The invention also provides vectors, and particularly expression vectors (*e.g.*, cos-Tet vector), which include any of the above-described nucleic acids. It is a further object of the invention to provide vectors in which normal or mutant PAMP nucleic acid sequences are operably joined to exogenous regulatory regions to produce altered patterns of expression, or to exogenous coding regions to produce fusion proteins. Conversely, it is another object to provide nucleic acids in which PAMP regulatory regions are operably joined to exogenous coding regions, including standard marker genes, to produce constructs in which the regulation of PAMP genes may be studied and used in assays or therapeutics.

The invention further provides host cells and transgenic animals transformed with any of the above-described nucleic acids of the invention. The host cells may be prokaryotic or eukaryotic cells and, in particular, may be gametes, zygotes, fetal cells, or stem cells useful in producing transgenic animal models. In one embodiment, the transgenic animal contains a transgene encoding a normal or mutant PAMP, which is expressed in neural cells such that expression can be detected, *e.g.*, by detecting PAMP, mRNA, or protein, and more preferably by detecting a neuroprotective or a neurodegenerative phenotype. For example, the animal might manifest one or more symptoms of a neurodegenerative disease. The animal may

be a vertebrate or an invertebrate. In a preferred embodiment, the transgenic animal is a mouse, which encodes a human PAMP. The transgenic animal may further comprise a second transgene encoding a normal or mutant PS1, PS2, or β APP.

In another embodiment, the invention provides an animal containing a nucleic acid that expresses a PAMP or a mutant PAMP at a higher or lower level relative to expression level in a wild-type animal. The animal may be prepared by homologous recombination mediated targeting of endogenous PAMP nucleic acid. In a preferred embodiment, the animal is prepared by translocation of P-elements or chemical mutagenesis.

The invention also provides a reconstituted system for measuring PAMP activity, comprising PAMP, a mutant PAMP, or functional fragments thereof, and a PAMP substrate. The reconstituted system may be a whole cell. Preferably, the whole cell contains a first nucleic acid that expresses said PAMP and a second nucleic acid that expresses the substrate. Preferably, the substrate comprises PS1 protein, PS2 protein, β APP, or a surrogate synthetic substrate protein such as Notch, which undergoes proteolytic processing events similar to those of β APP (Haass C and Selkoe DJ. *Nature* 1998; 391: 387-390; De Strooper B, *et al.*, *Nature* 1999; 398:518-522; Song W, *et al.*, *Proc Natl Acad Sci USA* 1999; 96: 6959-6963.; Struhl G and Greenwald I, *Nature* 1999; 398: 522-525; Ye Y, *et al.*, *Nature* 1999; 398: 525-529.).

The invention provides, in addition, a complex between a PAMP, or a mutant PAMP, and an agent which provides a detectable conformational or functional change in the PAMP upon interaction with a substance being analyzed for activity against a neurodegenerative disease. The complex may further comprise PS1 protein, PS2 protein or β APP.

The invention also provides a method for detecting a mutation in PAMP associated with Alzheimer's or a related neurological disorder, comprising obtaining a nucleic acid sample from an individual diagnosed with or suspected of having a neurodegenerative disorder, and sequencing a gene encoding PAMP from said sample.

The invention also provides a method for diagnosing individuals predisposed to or having a neurodegenerative disorder, comprising obtaining a nucleic acid sample from an individual diagnosed with or suspected of having a neurodegenerative disorder, and sequencing a gene encoding PAMP from said sample.

The invention also provides a method for diagnosing individuals predisposed to or having a neurodegenerative disorder, comprising obtaining cells that contain nucleic acid encoding PAMP, and under non-pathological conditions, transcribe the nucleic acid, and measuring a level of transcriptional activity of the nucleic acid.

The invention further provides a method for diagnosing individuals predisposed to or having a neurodegenerative disorder, comprising obtaining cells from an individual that express nucleic acid encoding PAMP, or isolating PAMP from said individual, and measuring PAMP activity, for example PAMP expression levels. In an alternative embodiment, the activity or abundance of a PAMP substrate may be measured.

The invention also provides a method for identifying putative agents having anti-neurodegenerative activity, comprising administering one or more putative agents to a transgenic animal and detecting a change in PAMP activity.

The invention also provides a method for identifying putative agents having anti-neurodegenerative activity, comprising adding one or more said agents to the reconstituted system described above, and detecting a change in PAMP activity.

The invention also provides a method for identifying putative agents having anti-neurodegenerative activity, comprising adding one or more said agents to the complex described above, and detecting a conformational change in PAMP.

The invention also provides a method for identifying proteins that interact with PAMP, comprising contacting said substance to the reconstituted system discussed above, and detecting a change in PAMP activity.

Further the invention provides for a method for identifying substances that modulate PAMP activity, comprising contacting a sample containing one or more substances with PAMP, or a PAMP mutant, or functional fragments thereof, and a PAMP substrate, measuring PAMP activity, and determining whether a change in PAMP activity occurs. In a preferred embodiment, the substance is a PAMP inhibitor. In another preferred embodiment, the substance stimulates PAMP activity.

These and other aspects of the invention are further elaborated in the Detailed Description of the Invention and Examples, *infra*.

DESCRIPTION OF THE DRAWINGS

FIGURE 1A and 1B. Predicted amino acid sequences for human (SEQ ID NO:14), mouse (SEQ ID NO:16), *D.melanogaster* (SEQ ID NO:18) and *C.elegans* (SEQ ID NO:12) orthologues. Residues not conserved in non-human PAMP are blank, conserved residues are aligned, similar residues are denoted by “+”.

DETAILED DESCRIPTION OF THE INVENTION

While PS1 and PS2 have been implicated in proper processing of β APP, and mutations in these proteins have been associated with Alzheimer's Disease, further

transmembrane receptor zinc glycoprotein (SWISS-PROT; World Wide Web (www) expasy.ch/sprot); Accession No. Q93332).

Table 1 - Potential functional sequence motifs in PAMP (SEQ ID NO: 14).

Potential function	PAMP Residue
N-asparaginyl glycosylation	44, 101, 290, 492, 698, 964, 1353, 1772, 2209, 2675, 3183, 3715, 4279, 4854, 5436, and 6050.
Glycosaminoglycan attachment	403
Myristolation	4, 37, 102, 226, 376, 548, 757, 1055, 1497, 1947, 2455, and 3035.
Phosphorylation sites for cAMP- and cGMP-dependent protein kinase	231
Phosphorylation sites for protein kinase C	114, 383, 724, 1109, 1499, 1983, 2598, and 3223
Phosphorylation sites for casein kinase II	7, 289, 652, 1026, 1483, 1951, 2425, 3068, and 3717

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The invention is further based on the identification of conserved functional domains, based on comparison and evaluation of the predicted amino acid sequences of human (SEQ ID NO: 14), murine (SEQ ID NO: 16), *D. melanogaster* (SEQ ID NO: 18), and *C. elegans* (SEQ ID NO: 12) orthologues of PAMP. "PAMP" can be characterized by the presence of conserved structural features, relative to orthologues from *D. melanogaster* and *C. elegans*. Nucleotide sequences encoding homologous hypothetical proteins exist in mice multiple EST, and *C. elegans* (GenBank; World Wide Web (www) ncbi.nlm.nih.gov; Accession No. Z75714; 37% similarity, $p = 8.7e^{-26}$) (Wilson *et al.*, *Nature* 1994; 368: 32-38). These hypothetical murine and nematode proteins have a similar topology and contain similar functional motifs to human PAMP. The existence of such homology predicts that similar proteins will be detected in other species including Xenopus, and Zebra fish, to mention a few such possibilities. By comparing the predicted amino acid sequences of human (SEQ ID NO: 14), murine (SEQ ID NO: 16), *D. melanogaster* (SEQ ID NO: 18), and *C. elegans* (SEQ ID NO: 12) PAMP proteins, we have deduced a series of conserved functional domains. One domain has chemical similarities to cyclic nucleotide binding domains of other proteins, and may have some regulatory role on a potential complex formed between PS1:PAMP and the C-terminal fragment of β APP, derived either from α - or β -secretase. These putative functional domains are sites for therapeutic target development by deploying drugs which might interact with these sites to modulate β APP processing via this complex.

The term "PAMP" also refers to functionally active fragments of the protein. Such fragments include, but are not limited to, peptides that contain an epitope, *e.g.*, as determined by conventional algorithms such as hydrophilicity/hydrophobicity analysis for antibody epitopes, and amphipathicity or consensus algorithms for T cell epitopes (Spouge, *et al.*, J. Immunol, 138:2204, 1987; Margalit, *et al.*, J. Immunol., 138:2213, 1987; Rothbard, Ann. Inst. Pateur., 137E:518, 1986; Rothbard and Taylor, EMBO J., 7:93, 1988). More preferably, a functionally active fragment of PAMP is a conserved domain, relative to the *D. melanogaster* and *C. elegans* orthologues. A specific functionally active fragment of PAMP is a fragment that interacts with PS1 or PS2, or both.

PAMP also encompasses naturally occurring variants, including other mammalian PAMPs (readily identified, as shown herein for murine PAMP, based on the presence of the structural features set forth above), allelic variants of PAMP from other human sources (including variants containing polymorphisms that are predictive of disease propensity or of response to pharmacological agents), and mutant forms of PAMP or *PAMP* genes that are associated with neurological diseases and disorders (such as spina bifida), particularly neurodegenerative diseases (such as AD). Also included are artificial PAMP mutants created by standard techniques such as site directed mutagenesis or chemical synthesis.

A PAMP "substrate" may be a polypeptide or protein, or any other type of compound, with which PAMP interacts physiologically. Examples of PAMP substrates include PS1, PS2, and β APP. Furthermore, A PAMP "ligand" may be a polypeptide, protein, lipid, carbohydrate, vitamin, mineral, amino acid, or any other type of compound which binds to PAMP. Hypothetically, PAMP may function as a receptor which modulates PS1/PS2/ β APP processing in response to signal (ligand) dependent interactions with PAMP.

Definitions

In accordance with the present invention there may be employed conventional molecular biology, microbiology, and recombinant DNA techniques within the skill of the art. Such techniques are explained fully in the literature. See, *e.g.*, Sambrook, Fritsch & Maniatis, *Molecular Cloning: A Laboratory Manual*, Second Edition (1989) Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York (herein "Sambrook *et al.*, 1989"); *DNA Cloning: A Practical Approach*, Volumes I and II (D.N. Glover ed. 1985); *Oligonucleotide Synthesis* (M.J. Gait ed. 1984); *Nucleic Acid Hybridization* (B.D. Hames & S.J. Higgins eds. (1985)); *Transcription And Translation* (B.D. Hames & S.J. Higgins, eds. (1984)); *Animal Cell Culture* (R.I. Freshney, ed. (1986)); *Immobilized Cells And Enzymes* (IRL Press, (1986)); B. Perbal, *A Practical Guide To Molecular Cloning* (1984); F.M. Ausubel *et al.* (eds.), *Current Protocols in Molecular Biology*, John Wiley & Sons, Inc. (1994).

If appearing herein, the following terms shall have the definitions set out below.

The use of italics (*e.g.*, *PAMP*) indicates a nucleic acid molecule (cDNA, mRNA, gene, etc.); normal text (*e.g.*, *PAMP*) indicates the polypeptide or protein.

In a specific embodiment, the term "about" or "approximately" means within 20%, preferably within 10%, and more preferably within 5% of a given value or range.

5 Alternatively, particularly in biological systems which are often responsive to order of magnitude changes, the term about means within an order of magnitude of a given value, preferably within a multiple of about 5-fold, and more preferably within a factor of about 2-fold of a given value.

As used herein, the term "isolated" means that the referenced material is free of
10 components found in the natural environment in which the material is normally found. In particular, isolated biological material is free of cellular components. In the case of nucleic acid molecules, an isolated nucleic acid includes a PCR product, an isolated mRNA, a cDNA, or a restriction fragment. In another embodiment, an isolated nucleic acid is preferably excised from the chromosome in which it may be found, and more preferably is no longer
15 joined to non-regulatory, non-coding regions, or to other genes, located upstream or downstream of the gene contained by the isolated nucleic acid molecule when found in the chromosome. In yet another embodiment, the isolated nucleic acid lacks one or more introns. Isolated nucleic acid molecules can be inserted into plasmids, cosmids, artificial chromosomes, and the like. Thus, in a specific embodiment, a recombinant nucleic acid is an isolated nucleic
20 acid. An isolated protein may be associated with other proteins or nucleic acids, or both, with which it associates in the cell, or with cellular membranes if it is a membrane-associated protein. An isolated organelle, cell, or tissue is removed from the anatomical site in which it is found in an organism. An isolated material may be, but need not be, purified.

The term "purified" as used herein refers to material that has been isolated
25 under conditions that reduce or eliminate unrelated materials, *i.e.*, contaminants. For example, a purified protein is preferably substantially free of other proteins or nucleic acids with which it is associated in a cell; a purified nucleic acid molecule is preferably substantially free of proteins or other unrelated nucleic acid molecules with which it can be found within a cell.

30 As used herein, the term "substantially free" is used operationally, in the context of analytical testing of the material. Preferably, purified material substantially free of contaminants is at least 50% pure; more preferably, at least 90% pure, and more preferably still at least 99% pure. Purity can be evaluated by chromatography, gel electrophoresis, immunoassay, composition analysis, biological assay, and other methods known in the art.

35 The term "host cell" means any cell of any organism that is selected, modified, transformed, grown, or used or manipulated in any way, for the production of a substance by

the cell, for example the expression by the cell of a gene, a DNA or RNA sequence, a protein or an enzyme. Host cells can further be used for screening or functional assays, as described *infra*. A host cell has been "transfected" by exogenous or heterologous DNA when such DNA has been introduced inside the cell. A cell has been "transformed" by exogenous or heterologous DNA when the transfected DNA is expressed and effects a function or phenotype on the cell in which it is expressed. The term "expression system" means a host cell transformed by a compatible expression vector and cultured under suitable conditions *e.g.* for the expression of a protein coded for by foreign DNA carried by the vector and introduced to the host cell.

Proteins and polypeptides can be made in the host cell by expression of recombinant DNA. As used herein, the term "polypeptide" refers to an amino acid-based polymer, which can be encoded by a nucleic acid or prepared synthetically. Polypeptides can be proteins, protein fragments, chimeric proteins, etc. Generally, the term "protein" refers to a polypeptide expressed endogenously in a cell, *e.g.*, the naturally occurring form (or forms) of the amino acid-based polymer.

A "coding sequence" or a sequence "encoding" an expression product, such as a RNA, polypeptide, protein, or enzyme, is a nucleotide sequence that, when expressed, results in the production of that RNA, polypeptide, protein, or enzyme, *i.e.*, the nucleotide sequence encodes an amino acid sequence for that polypeptide, protein or enzyme. A coding sequence for a protein may include a start codon (usually ATG) and a stop codon.

The coding sequences herein may be flanked by natural regulatory (expression control) sequences, or may be associated with heterologous sequences, including promoters, internal ribosome entry sites (IRES) and other ribosome binding site sequences, enhancers, response elements, suppressors, signal sequences, polyadenylation sequences, introns, 5'- and 3'- non-coding regions, and the like. The nucleic acids may also be modified by many means known in the art. Non-limiting examples of such modifications include methylation, "caps", substitution of one or more of the naturally occurring nucleotides with an analog, and internucleotide modifications.

The term "gene", also called a "structural gene" means a DNA sequence that codes for or corresponds to a particular sequence of ribonucleic acids or amino acids which comprise all or part of one or more proteins, and may or may not include regulatory DNA sequences, such as promoter sequences, which determine for example the conditions under which the gene is expressed.

A "promoter sequence" is a DNA regulatory region capable of binding RNA polymerase in a cell and initiating transcription of a downstream (3' direction) coding sequence. For purposes of defining the present invention, the promoter sequence is bounded at

its 3' terminus by the transcription initiation site and extends upstream (5' direction) to include the minimum number of bases or elements necessary to initiate transcription at levels detectable above background.

A coding sequence is "under the control" or "operatively associated with" of transcriptional and translational control sequences in a cell when RNA polymerase transcribes the coding sequence into mRNA, which then may be trans-RNA spliced (if it contains introns) and translated into the protein encoded by the coding sequence.

The terms "express" and "expression" mean allowing or causing the information in a gene or DNA sequence to become manifest, for example producing a protein by activating the cellular functions involved in transcription and translation of a corresponding gene or DNA sequence. A DNA sequence is expressed in or by a cell to form an "expression product" such as a protein. The expression product itself, *e.g.* the resulting protein, may also be said to be "expressed" by the cell.

The term "transfection" means the introduction of a foreign nucleic acid into a cell. The term "transformation" means the introduction of a "foreign" (*i.e.* extrinsic or extracellular) gene, DNA or RNA sequence to a host cell, so that the host cell will express the introduced gene or sequence to produce a desired substance, typically a protein or enzyme coded by the introduced gene or sequence. The introduced gene or sequence may also be called a "cloned", "foreign", or "heterologous" gene or sequence, and may include regulatory or control sequences used by a cell's genetic machinery. The gene or sequence may include nonfunctional sequences or sequences with no known function. A host cell that receives and expresses introduced DNA or RNA has been "transformed" and is a "transformant" or a "clone." The DNA or RNA introduced to a host cell can come from any source, including cells of the same genus or species as the host cell, or cells of a different genus or species.

The terms "vector", "cloning vector" and "expression vector" mean the vehicle by which a DNA or RNA sequence (*e.g.*, a foreign gene) can be introduced into a host cell, so as to transform the host and promote expression (*e.g.*, transcription and translation) of the introduced sequence. Vectors include plasmids, phages, viruses, etc. A "cassette" refers to a DNA coding sequence or segment of DNA that codes for an expression product that can be inserted into a vector at defined restriction sites. The cassette restriction sites are designed to ensure insertion of the cassette in the proper reading frame. Generally, foreign DNA is inserted at one or more restriction sites of the vector DNA, and then is carried by the vector into a host cell along with the transmissible vector DNA. A segment or sequence of DNA having inserted or added DNA, such as an expression vector, can also be called a "DNA construct." Recombinant cloning vectors will often include one or more replication systems

for cloning or expression, one or more markers for selection in the host, *e.g.* antibiotic resistance, and one or more expression cassettes.

A "knockout mammal" is a mammal (*e.g.*, mouse) that contains within its genome a specific gene that has been inactivated by the method of gene targeting (*see, e.g.*, US Patents No. 5,777,195 and No. 5,616,491). A knockout mammal includes both a heterozygote knockout (*i.e.*, one defective allele and one wild-type allele) and a homozygous mutant. Preparation of a knockout mammal requires first introducing a nucleic acid construct that will be used to suppress expression of a particular gene into an undifferentiated cell type termed an embryonic stem cell. This cell is then injected into a mammalian embryo. A mammalian embryo with an integrated cell is then implanted into a foster mother for the duration of gestation. Zhou, *et al.* (Genes and Development, 9:2623-34, 1995) describes PPCA knock-out mice. Knockout mice can be used to study defects in neurological development or neurodegenerative diseases. Disease phenotypes that develop can provide a platform for further drug discovery.

The term "knockout" refers to partial or complete suppression of the expression of at least a portion of a protein encoded by an endogenous DNA sequence in a cell. The term "knockout construct" refers to a nucleic acid sequence that is designed to decrease or suppress expression of a protein encoded by endogenous DNA sequences in a cell. The nucleic acid sequence used as the knockout construct is typically comprised of (1) DNA from some portion of the gene (exon sequence, intron sequence, and/or promoter sequence) to be suppressed and (2) a marker sequence used to detect the presence of the knockout construct in the cell. The knockout construct is inserted into a cell, and integrates with the genomic DNA of the cell in such a position so as to prevent or interrupt transcription of the native DNA sequence. Such insertion usually occurs by homologous recombination (*i.e.*, regions of the knockout construct that are homologous to endogenous DNA sequences hybridize to each other when the knockout construct is inserted into the cell and recombine so that the knockout construct is incorporated into the corresponding position of the endogenous DNA). The knockout construct nucleic acid sequence may comprise 1) a full or partial sequence of one or more exons and/or introns of the gene to be suppressed, 2) a full or partial promoter sequence of the gene to be suppressed, or 3) combinations thereof. Typically, the knockout construct is inserted into an embryonic stem cell (ES cell) and is integrated into the ES cell genomic DNA, usually by the process of homologous recombination. This ES cell is then injected into, and integrates with, the developing embryo.

Generally, for homologous recombination, the DNA will be at least about 1 kilobase (kb) in length and preferably 3-4 kb in length, thereby providing sufficient

complementary sequence for recombination when the knockout construct is introduced into the genomic DNA of the ES cell.

A "knock-in" mammal is a mammal in which an endogenous gene is substituted with a heterologous gene or a modified variant of the endogenous gene (Roemer *et al.*, New Biol. 3:331, 1991). Preferably, the heterologous gene is "knocked-in" to a locus of interest, for example into a gene that is the subject of evaluation of expression or function, thereby linking the heterologous gene expression to transcription from the appropriate promoter (in which case the gene may be a reporter gene; *see* Elefanty *et al.*, Proc Natl Acad Sci USA 95:11897, 1998). This can be achieved by homologous recombination, transposon (Westphal and Leder, Curr Biol 7:530, 1997), using mutant recombination sites (Araki *et al.*, Nucleic Acids Res 25:868, 1997) or PCR (Zhang and Henderson, Biotechniques 25:784, 1998).

The phrases "disruption of the gene" and "gene disruption" refer to insertion of a nucleic acid sequence into one region of the native DNA sequence (usually one or more exons) and/or the promoter region of a gene so as to decrease or prevent expression of that gene in the cell as compared to the wild-type or naturally occurring sequence of the gene. By way of example, a nucleic acid construct can be prepared containing a DNA sequence encoding an antibiotic resistance gene which is inserted into the DNA sequence that is complementary to the DNA sequence (promoter and/or coding region) to be disrupted. When this nucleic acid construct is then transfected into a cell, the construct will integrate into the genomic DNA. Thus, some progeny of the cell will no longer express the gene, or will express it at a decreased level, as the DNA is now disrupted by the antibiotic resistance gene.

The term "heterologous" refers to a combination of elements not naturally occurring. For example, heterologous DNA refers to DNA not naturally located in the cell, or in a chromosomal site of the cell. Preferably, the heterologous DNA includes a gene foreign to the cell. A heterologous expression regulatory element is a such an element operatively associated with a different gene than the one it is operatively associated with in nature. In the context of the present invention, an gene is heterologous to the recombinant vector DNA in which it is inserted for cloning or expression, and it is heterologous to a host cell containing such a vector, in which it is expressed, *e.g.*, a CHO cell.

The terms "mutant" and "mutation" mean any detectable change in genetic material, *e.g.* DNA, or any process, mechanism, or result of such a change. This includes gene mutations, in which the structure (*e.g.*, DNA sequence) of a gene is altered, any gene or DNA arising from any mutation process, and any expression product (*e.g.*, protein) expressed by a modified gene or DNA sequence. The term "variant" may also be used to indicate a modified or altered gene, DNA sequence, enzyme, cell, etc., *i.e.*, any kind of mutant.

"Sequence-conservative variants" of a polynucleotide sequence are those in which a change of one or more nucleotides in a given codon position results in no alteration in the amino acid encoded at that position.

"Function-conservative variants" are those in which a given amino acid residue in a protein or enzyme has been changed without altering the overall conformation and function of the polypeptide, including, but not limited to, replacement of an amino acid with one having similar properties (such as, for example, polarity, hydrogen bonding potential, acidic, basic, hydrophobic, aromatic, and the like). Amino acids with similar properties are well known in the art. For example, arginine, histidine and lysine are hydrophilic-basic amino acids and may be interchangeable. Similarly, isoleucine, a hydrophobic amino acid, may be replaced with leucine, methionine or valine. Such changes are expected to have little or no effect on the apparent molecular weight or isoelectric point of the protein or polypeptide. Amino acids other than those indicated as conserved may differ in a protein or enzyme so that the percent protein or amino acid sequence similarity between any two proteins of similar function may vary and may be, for example, from 70% to 99% as determined according to an alignment scheme such as by the Cluster Method, wherein similarity is based on the MEGALIGN algorithm. A "function-conservative variant" also includes a polypeptide or enzyme which has at least 60% amino acid identity as determined by BLAST (Altschul SF, *et al.*, J Mol Biol 1990; 215: 403-410) or FASTA algorithms, preferably at least 75%, most preferably at least 85%, and even more preferably at least 90%, and which has the same or substantially similar properties or functions as the native or parent protein or enzyme to which it is compared.

An "ortholog" to a protein means a corresponding protein from another species. Orthologous proteins typically have similar functions in different species, and can also be substantially homologous.

As used herein, the term "homologous" in all its grammatical forms and spelling variations refers to the relationship between proteins that possess a "common evolutionary origin," including proteins from superfamilies (*e.g.*, the immunoglobulin superfamily) and homologous proteins from different species (*e.g.*, myosin light chain, etc.) (Reeck *et al.*, Cell 50:667, 1987). Such proteins (and their encoding genes) have sequence homology, as reflected by their sequence similarity, whether in terms of percent similarity or the presence of specific residues or motifs. Motif analysis can be performed using, for example, the program BLOCKS (World Wide Web (www) blocks.fhcrc.org). Accordingly, the term "sequence similarity" in all its grammatical forms refers to the degree of identity or correspondence between nucleic acid or amino acid sequences of proteins that may or may not share a common evolutionary origin (*see* Reeck *et al.*, *supra*). However, in common usage and in the instant application, the

term "homologous," when modified with an adverb such as "highly," may refer to sequence similarity and may or may not relate to a common evolutionary origin.

In a specific embodiment, two DNA sequences are "substantially homologous" or "substantially similar" when at least about 80%, and most preferably at least about 90 or 95% of the nucleotides match over the defined length of the DNA sequences, as determined by sequence comparison algorithms, such as BLAST, FASTA, DNA Strider, etc. Sequences that are substantially homologous can be identified by comparing the sequences using standard software available in sequence data banks, or in a Southern hybridization experiment under, for example, stringent conditions as defined for that particular system.

Similarly, in a particular embodiment, two amino acid sequences are "substantially homologous" or "substantially similar" when greater than 80% of the amino acids are identical, or greater than about 90% are similar (functionally identical). Preferably, the similar or homologous sequences are identified by alignment using, for example, the GCG (Genetics Computer Group, Program Manual for the GCG Package, Version 7, Madison, Wisconsin) pileup program, ProteinPredict (dodo.cmpc.columbia.edu/predictprotein), or any of the programs described above (BLAST, FASTA, etc.).

A nucleic acid molecule is "hybridizable" to another nucleic acid molecule, such as a cDNA, genomic DNA, or RNA, when a single stranded form of the nucleic acid molecule can anneal to the other nucleic acid molecule under the appropriate conditions of temperature and solution ionic strength (*see* Sambrook *et al.*, *supra*). The conditions of temperature and ionic strength determine the "stringency" of the hybridization. For preliminary screening for homologous nucleic acids, low stringency hybridization conditions, corresponding to a T_m (melting temperature) of 55°C, can be used. Moderate stringency hybridization conditions correspond to a higher T_m and high stringency hybridization conditions correspond to the highest T_m . Hybridization requires that the two nucleic acids contain complementary sequences, although depending on the stringency of the hybridization, mismatches between bases are possible. The appropriate stringency for hybridizing nucleic acids depends on the length of the nucleic acids and the degree of complementation, variables well known in the art. The greater the degree of similarity or homology between two nucleotide sequences, the greater the value of T_m for hybrids of nucleic acids having those sequences. The relative stability (corresponding to higher T_m) of nucleic acid hybridizations decreases in the following order: RNA:RNA, DNA:RNA, DNA:DNA. For hybrids of greater than 100 nucleotides in length, equations for calculating T_m have been derived (*see* Sambrook *et al.*, *supra*, 9.50-9.51). For hybridization with shorter nucleic acids, *i.e.*, oligonucleotides, the position of mismatches becomes more important, and the length of the oligonucleotide determines its specificity (*see* Sambrook *et al.*, *supra*, 11.7-11.8). A minimum length for a hybridizable

nucleic acid is at least about 10 nucleotides; preferably at least about 15 nucleotides; and more preferably the length is at least about 20 nucleotides.

The present invention provides antisense nucleic acids (including ribozymes), which may be used to inhibit expression of PAMP,, *e.g.*, to disrupt a cellular process (such
5 disruption can be used in an animal model or therapeutically). An "antisense nucleic acid" is a single stranded nucleic acid molecule which, on hybridizing under cytoplasmic conditions with complementary bases in an RNA or DNA molecule, inhibits the latter's role. If the RNA is a messenger RNA transcript, the antisense nucleic acid is a countertranscript or mRNA-interfering complementary nucleic acid. As presently used, "antisense" broadly includes RNA-
10 RNA interactions, RNA-DNA interactions, ribozymes and RNase-H mediated arrest. Antisense nucleic acid molecules can be encoded by a recombinant gene for expression in a cell (*e.g.*, U.S. Patent No. 5,814,500; U.S. Patent No. 5,811,234), or alternatively they can be prepared synthetically (*e.g.*, U.S. Patent No. 5,780,607).

As used herein, the term "oligonucleotide" refers to a nucleic acid, generally of
15 at least 10, preferably at least 15, and more preferably at least 20 nucleotides, preferably no more than 100 nucleotides, that is hybridizable to a genomic DNA molecule, a cDNA molecule, or an mRNA molecule encoding a gene, mRNA, cDNA, or other nucleic acid of interest. Oligonucleotides can be labeled, *e.g.*, with ³²P-nucleotides or nucleotides to which a label, such as biotin, has been covalently conjugated. In one embodiment, a labeled
20 oligonucleotide can be used as a probe to detect the presence of a nucleic acid. In another embodiment, oligonucleotides (one or both of which may be labeled) can be used as PCR primers, *e.g.*, for cloning full length or a fragment of a protein or polypeptide. In a further embodiment, an oligonucleotide of the invention can form a triple helix with a nucleic acid (genomic DNA or mRNA) encoding a protein or polypeptide. Generally, oligonucleotides are
25 prepared synthetically, preferably on a nucleic acid synthesizer. Accordingly, oligonucleotides can be prepared with non-naturally occurring phosphoester analog bonds, such as thioester bonds, etc. Furthermore, the oligonucleotides herein may also be modified with a label capable of providing a detectable signal, either directly or indirectly. Exemplary labels include radioisotopes, fluorescent molecules, biotin, and the like.

30 Specific non-limiting examples of synthetic oligonucleotides envisioned for this invention include oligonucleotides that contain phosphorothioates, phosphotriesters, methyl phosphonates, short chain alkyl, or cycloalkyl intersugar linkages or short chain heteroatomic or heterocyclic intersugar linkages. Most preferred are those with CH₂-NH-O-CH₂, CH₂-N(CH₃)-O-CH₂, CH₂-O-N(CH₃)-CH₂, CH₂-N(CH₃)-N(CH₃)-CH₂ and O-N(CH₃)-CH₂-CH₂ backbones
35 (where phosphodiester is O-PO₂-O-CH₂). US Patent No. 5,677,437 describes heteroaromatic oligonucleoside linkages. Nitrogen linkers or groups containing nitrogen can also be used to

prepare oligonucleotide mimics (U.S. Patents No. 5,792,844 and No. 5,783,682). US Patent No. 5,637,684 describes phosphoramidate and phosphorothioamidate oligomeric compounds. Also envisioned are oligonucleotides having morpholino backbone structures (U.S. Pat. No. 5,034,506). In other embodiments, such as the peptide-nucleic acid (PNA) backbone, the phosphodiester backbone of the oligonucleotide may be replaced with a polyamide backbone, the bases being bound directly or indirectly to the aza nitrogen atoms of the polyamide backbone (Nielsen *et al.*, Science 254:1497, 1991). Other synthetic oligonucleotides may contain substituted sugar moieties comprising one of the following at the 2' position: OH, SH, SCH₃, F, OCN, O(CH₂)_nNH₂ or O(CH₂)_nCH₃ where n is from 1 to about 10; C₁ to C₁₀ lower alkyl, substituted lower alkyl, alkaryl or aralkyl; Cl; Br; CN; CF₃; OCF₃; O-, S-, or N-alkyl; O-, S-, or N-alkenyl; SOCH₃; SO₂CH₃; ONO₂; NO₂; N₃; NH₂; heterocycloalkyl; heterocycloalkaryl; aminoalkylamino; polyalkylamino; substituted silyl; a fluorescein moiety; an RNA cleaving group; a reporter group; an intercalator; a group for improving the pharmacokinetic properties of an oligonucleotide; or a group for improving the pharmacodynamic properties of an oligonucleotide, and other substituents having similar properties. Oligonucleotides may also have sugar mimetics such as cyclobutyls or other carbocyclics in place of the pentofuranosyl group. Nucleotide units having nucleosides other than adenosine, cytidine, guanosine, thymidine and uridine, such as inosine, may be used in an oligonucleotide molecule.

Presenilin interacting proteins

Mutant PS1 and PS2 genes, and their corresponding amino acid sequences are described in Applicants' co-pending U.S. Application Serial No. 08/888,077, filed July 3, 1997, and incorporated herein by reference. Examples of PS1 mutations include I143T, M146L, L171P, F177S, A260V, C263R, P264L, P267S, E280A, E280G, A285V, L286V, Δ291-319, L322V, G384A, L392V, C410Y and I439V. Examples of PS2 mutations include N141I, M239V and I420T.

The methods of the present invention are not limited to mutant presenilins wherein the PAMP-interacting domain is mutated relative to the wild-type protein. For example, Applicants have observed that mutations in PS1 (*e.g.*, M146L) outside of the interacting domain (loop) also affect β-catenin translocation. These mutations probably disturb the presenilin *armadillo* interactions by altering the function of a high MW complex which contains, *e.g.*, the presenilin and *armadillo* proteins, as described in Yu *et al.*, 1998, *J. Biol. Chem.* 273:16460-16475. Moreover, a comparison of the human PS1 (hPS1) and PS2 (hPS2) sequences reveals that these pathogenic mutations are in regions of the PS1 protein which are conserved in the PS2 protein. Therefore, corresponding mutations in corresponding regions of PS2 may also be expected to be pathogenic and are useful in the methods described herein.

Proteins that interact with the presenilins, *i.e.*, PS-interacting proteins, include PAMP, the S5a subunit of the 26S proteasome (GenBank; Accession No. U51007), Rab11 (GenBank; Accession Nos. X56740 and X53143), retinoid X receptor B, also known as nuclear receptor co-regulator or MHC (GenBank Accession Nos. M84820, and X63522), GT24 (GenBank Accession No. U81004), β -catenin (*Zhou et al.*, 1997, *Neuro. Report* (Fast Track) 8:1025-1029 and *Yu et al.*, *supra*) as well as *armadillo* proteins. These and other PS1 binding proteins are described in Applicants' copending commonly assigned U.S. Application Serial No. 08/888,077, filed July 3, 1997, as well as U.S. Application Serial No. 08/592,541, filed January 26, 1996, the disclosures of which are incorporated herein by reference.

PAMP Mutants

PAMP mutants may cause biochemical changes similar to those affecting the onset or progression of Alzheimer Disease. Therefore, artificial PAMP mutations can potentially be used to generate cellular and other model systems to design treatments and preventions for Alzheimer Disease related disorders. Such mutations may also be used for evaluating whether PAMP is involved in the pathogenesis of AD. Since the amyloid- β (A β) inducing mutations are found in amino acid residues of a soluble (non-membrane spanning) domain of PAMP, analysis of the normal structure of this domain and the effects of these and other nearby mutations on the structure of this domain (and the other domains of PAMP) provide information for the design of specific molecular therapeutics.

In general, modifications of the sequences encoding the polypeptides described herein may be readily accomplished by standard techniques such as chemical syntheses and site-directed mutagenesis. See *Gillman et al.*, 1979, *Gene* 8:81-97; *Roberts et al.*, 1987, *Nature* 328:731-734; and *Innis* (ed.), 1990, *PCR Protocols: A Guide to Methods and Applications*, Academic Press, New York. Most modifications are evaluated by routine screening via an assay designed to select for the desired property.

Antibodies to PAMP

According to the invention, PAMP polypeptides produced recombinantly or by chemical synthesis, and fragments or other derivatives or analogs thereof, including fusion proteins and PAMP mutants, may be used as an immunogen to generate antibodies that recognize the PAMP polypeptide. Such antibodies include but are not limited to polyclonal, monoclonal, chimeric, single chain, Fab fragments, and an Fab expression library. Such an antibody is preferably specific for human PAMP, PAMP originating from other species, or for post-translationally modified (e.g. phosphorylated, glycosylated) PAMP.

Various procedures known in the art may be used for the production of polyclonal antibodies to PAMP polypeptide or derivative or analog thereof. For the production of antibody, various host animals can be immunized by injection with the PAMP polypeptide,

or a derivative (*e.g.*, fragment or fusion protein) thereof, including but not limited to rabbits, mice, rats, sheep, goats, etc. In one embodiment, the PAMP polypeptide or fragment thereof can be conjugated to an immunogenic carrier, *e.g.*, bovine serum albumin (BSA) or keyhole limpet hemocyanin (KLH). Various adjuvants may be used to increase the immunological response, depending on the host species, including but not limited to Freund's (complete and incomplete), mineral gels such as aluminum hydroxide, surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, keyhole limpet hemocyanins, dinitrophenol, and potentially useful human adjuvants such as BCG (*bacille Calmette-Guerin*) and *Corynebacterium parvum*. Antisera may be collected at a chosen time point after immunization, and purified as desired.

For preparation of monoclonal antibodies directed toward the PAMP polypeptide, or fragment, analog, or derivative thereof, any technique that provides for the production of antibody molecules by continuous cell lines in culture may be used. These include but are not limited to the hybridoma technique originally developed by Kohler and Milstein (Nature 256:495-497, 1975), as well as the trioma technique, the human B-cell hybridoma technique (Kozbor *et al.*, Immunology Today 4:72, 1983; Cote *et al.*, Proc. Natl. Acad. Sci. U.S.A. 80:2026-2030, 1983), and the EBV-hybridoma technique to produce human monoclonal antibodies (Cole *et al.*, in Monoclonal Antibodies and Cancer Therapy, Alan R. Liss, Inc., pp. 77-96, 1985). Production of human antibodies by CDR grafting is described in U.S. Patent Nos. 5,585,089, 5,693,761, and 5,693,762 to Queen *et al.*, and also in U.S. Patent No. 5,225,539 to Winter and International Patent Application PCT/WO91/09967 by Adau *et al.* In an additional embodiment of the invention, monoclonal antibodies can be produced in germ-free animals (International Patent Publication No. WO 89/12690, published 28 December 1989). In fact, according to the invention, techniques developed for the production of "chimeric antibodies" (Morrison *et al.*, J. Bacteriol. 159:870, 1984; Neuberger *et al.*, Nature 312:604-608, 1984; Takeda *et al.*, Nature 314:452-454, 1985) by splicing the genes from a mouse antibody molecule specific for an PAMP polypeptide together with genes from a human antibody molecule of appropriate biological activity can be used; such antibodies are within the scope of this invention. Such human or humanized chimeric antibodies are preferred for use in therapy of human diseases or disorders (described *infra*), since the human or humanized antibodies are much less likely than xenogenic antibodies to induce an immune response, in particular an allergic response, themselves.

According to the invention, techniques described for the production of single chain antibodies (U.S. Patent Nos. 5,476,786 and 5,132,405 to Huston; U.S. Patent 4,946,778) can be adapted to produce PAMP polypeptide-specific single chain antibodies. An additional embodiment of the invention utilizes the techniques described for the construction of

Fab expression libraries (Huse *et al.*, Science 246:1275-1281, 1989) to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity for an PAMP polypeptide, or its derivatives, or analogs.

Antibody fragments which contain the idiotype of the antibody molecule can be generated by known techniques. For example, such fragments include but are not limited to: the F(ab')₂ fragment which can be produced by pepsin digestion of the antibody molecule; the Fab' fragments which can be generated by reducing the disulfide bridges of the F(ab')₂ fragment, and the Fab fragments which can be generated by treating the antibody molecule with papain and a reducing agent.

In the production of antibodies, screening for the desired antibody can be accomplished by techniques known in the art, *e.g.*, radioimmunoassay, ELISA (enzyme-linked immunosorbent assay), "sandwich" immunoassays, immunoradiometric assays, gel diffusion precipitin reactions, immunodiffusion assays, *in situ* immunoassays (using colloidal gold, enzyme or radioisotope labels, for example), western blots, precipitation reactions, agglutination assays (*e.g.*, gel agglutination assays, hemagglutination assays), complement fixation assays, immunofluorescence assays, protein A assays, and immunoelectrophoresis assays, etc. In one embodiment, antibody binding is detected by detecting a label on the primary antibody. In another embodiment, the primary antibody is detected by detecting binding of a secondary antibody or reagent to the primary antibody. In a further embodiment, the secondary antibody is labeled. Many means are known in the art for detecting binding in an immunoassay and are within the scope of the present invention. For example, to select antibodies which recognize a specific epitope of an PAMP polypeptide, one may assay generated hybridomas for a product which binds to an PAMP polypeptide fragment containing such epitope. For selection of an antibody specific to an PAMP polypeptide from a particular species of animal, one can select on the basis of positive binding with PAMP polypeptide expressed by or isolated from cells of that species of animal.

The foregoing antibodies can be used in methods known in the art relating to the localization and activity of the PAMP polypeptide, *e.g.*, for Western blotting, imaging PAMP polypeptide *in situ*, measuring levels thereof in appropriate physiological samples, etc. using any of the detection techniques mentioned above or known in the art. Such antibodies can be used to identify proteins that interact with PAMP, and to detect conformational or structural changes in PAMP.

In a specific embodiment, antibodies that agonize or antagonize the activity of PAMP polypeptide can be generated. They can also be used to regulate or inhibit PAMP activity intracellular, *i.e.*, the invention contemplates an intracellular antibody (intrabody), *e.g.*, single chain Fv antibodies (*see generally*, Chen, Mol. Med. Today, 3:160-167, 1997;

Spitz *et al.*, Anticancer Res., 16:3415-3422, 1996; Indolfi *et al.*, Nat. Med., 2:634-635, 1996; Kijima *et al.*, Pharmacol. Ther., 68:247-267, 1995).

PAMP Diagnostic Assays

The nucleotide sequence and the protein sequence and the putative biological activity of PAMP or PAMP mutants can all be used for the purposes of diagnosis of individuals who are at-risk for, or who actually have, a variety of neurodegenerative diseases (including Alzheimer's disease, Lewy body variant, Parkinson's disease-dementia complex, amyotrophic lateral sclerosis etc.), neuropsychiatric diseases (schizophrenia, depression, mild cognitive impairment, benign senescent forgetfulness, age-associated memory loss, etc.), developmental disorders associated with defects in intracellular signal transduction mediated by factors such as Notch, Delta, Wingless, etc., and neoplasms (*e.g.*, bowel cancer, etc.) associated with abnormalities of PS1/PAMP/PS2 mediated regulation of cell death pathways. These diagnostic entities can be used by searching for alterations in: the nucleotide sequence of PAMP; in the transcriptional activity of PAMP; in the protein level as monitored by immunological means (*e.g.*, ELISA and Western blots); in the amino acid sequence (as ascertained by Western blotting, amino acid sequence analysis, mass spectroscopy); and/or in the biological activity of the PAMP protein as measured by either *in vivo* methods (*e.g.*, monitoring β APP processing and the production of amyloid- β peptide ($A\beta$), or other suitable protein substrates for PAMP including Notch, etc.), or by *in vitro* assays (using either whole cell or cell-free assays to measure processing of suitable substrates including β APP or parts thereof, and other proteins such as Notch). Any of these assays can also be performed in a transgenic animal model as well, *e.g.*, to measure the effect of a drug or a mutation or overexpression of a different gene *in vivo*.

PAMP Screening Assays

Identification and isolation of PAMP provides for development of screening assays, particularly for high throughput screening of molecules that up- or down-regulate the activity of PAMP, *e.g.*, by permitting expression of PAMP in quantities greater than can be isolated from natural sources, or in indicator cells that are specially engineered to indicate the activity of PAMP expressed after transfection or transformation of the cells. Any screening technique known in the art can be used to screen for PAMP agonists or antagonists. The present invention contemplates screens for small molecule ligands or ligand analogs and mimics, as well as screens for natural ligands that bind to and agonize or antagonize the activity of PAMP *in vivo*. For example, natural products libraries can be screened using assays of the invention for molecules that agonize or antagonize PAMP activity.

Another approach uses recombinant bacteriophage to produce large libraries. Using the "phage method" (Scott and Smith, Science 249:386-390, 1990; Cwirla, *et al.*, Proc.

Natl. Acad. Sci., 87:6378-6382, 1990; Devlin *et al.*, Science, 49:404-406, 1990), very large libraries can be constructed (10^6 - 10^8 chemical entities). A second approach uses primarily chemical methods, of which the Geysen method (Geysen *et al.*, Molecular Immunology 23:709-715, 1986; Geysen *et al.* J. Immunologic Method 102:259-274, 1987; and the method of Fodor *et al.* (Science 251:767-773, 1991) are examples. Furka *et al.* (14th Int. Congress of Biochemistry, Volume 5, Abstract FR:013, 1988; Furka, Int. J. Peptide Protein Res. 37:487-493, 1991), Houghton (U.S. Patent No. 4,631,211, issued December 1986) and Rutter *et al.* (U.S. Patent No. 5,010,175, issued April 23, 1991) describe methods to produce a mixture of peptides that can be tested as agonists or antagonists.

In another aspect, synthetic libraries (Needels *et al.*, Proc. Natl. Acad. Sci. USA 90:10700-4, 1993; Ohlmeyer *et al.*, Proc. Natl. Acad. Sci. USA 90:10922-10926, 1993; Lam *et al.*, International Patent Publ. No. WO 92/00252; Kocis *et al.*, International Patent Publ. No. WO 9428028) and the like can be used to screen for PAMP ligands according to the present invention.

Knowledge of the primary sequence of the protein, and the similarity of that sequence with proteins of known function, can provide an initial clue as to the inhibitors or antagonists of the protein. As noted above, identification and screening of antagonists is further facilitated by determining structural features of the protein, *e.g.*, using X-ray crystallography, neutron diffraction, nuclear magnetic resonance spectrometry, and other techniques for structure determination. These techniques provide for the rational design or identification of agonists and antagonists.

The PAMP protein sequence (including parts thereof) can be used to deduce the structural organization and topology of PAMP through the use of a variety of techniques including CD spectroscopy, nuclear magnetic resonance (NMR) spectroscopy, X-ray crystallography, and molecular modeling. Sequences for PAMP or PAMP mutants can also be used to identify proteins which interact with PAMP either in concert with PS1 and PS2, or independently, using a variety of methods including co-immunoprecipitation, yeast two hybrid interaction trap assays, yeast three hybrid interaction trap assays, chemical cross-linking and co-precipitation studies, etc. These and other methods are described more fully in co-pending and commonly assigned U.S. Application Serial No. 08/888,077, filed July 3, 1997, and 09/227,725, filed January 8, 1999, both of which are incorporated herein by reference. Identification of these interacting partners will then lead to their use to further delineate the biochemical pathways leading to the above-mentioned diseases.

Finally, the structural analysis of PAMP, when combined with structural analysis of PS1 and PS2, and other proteins which interact with PAMP or PAMP mutants, will identify the structural domains that mediate interactions between these molecules and which

also confer biological activity on PAMP (or PAMP and these other molecules). These structural domains, and other functional domains, which can modulate the activity of these structural domains, can all be modified through a variety of means, including but not limited to site-directed mutagenesis, in order to either augment or reduce the biological activity. The structure and topology of these domains can all be used as a basis for the rational design of pharmaceuticals (small molecule conventional drugs or novel carbohydrate, lipid, DNA/RNA or protein-based high molecular weight biological compounds) to modulate (increase or decrease) the activity of PAMP and/or the PAMP PS1/PS2 complex, and/or the activity of the PAMP/other protein complexes. For example, using structural prediction calculations, possibly in conjunction with spectroscopic data like nuclear magnetic resonance, circular dichroism, and other physical-chemical structural data, or crystallographic data, or both, one can generate molecular models for the structure of PAMP. These models, in turn, are important for rational drug design. Drug candidates generated using a rational drug design program can then be applied for the treatment and/or prevention of the above-mentioned diseases, and can be administered through a variety of means including: as conventional small molecules through enteral or parenteral routes; via inclusion in liposome vehicles; through infusion in pumps inserted into various organs (*e.g.*, via Omay pumps inserted into the cerebral ventricles); via the transplantation of genetically-modified cells expressing recombinant genes; or via the use of biological vectors (*e.g.*, retrovirus, adenovirus, adeno-associated virus, Lentivirus, or herpes simplex virus-based vectors) which allow targeted expression of appropriately modified gene products in selected cell types. It should be noted that the recombinant proteins described above may be the wild-type PAMP, a genetically-modified PAMP, a wild-type PS1/PS2, a genetically-modified PS1/PS2, or a specially-designed protein or peptide which is designed to interact with either the functional domains of PAMP (or the PAMP/PS1/PS2/other protein complex) or to interact with the domains which modulate the activity of the functional domains of PAMP.

PAMP *In Vitro* and *In Vivo* Models

The PAMP nucleotide sequence can be used to make cell-free systems, transfected cell lines, and animal models (invertebrate or vertebrate) of the neurodegenerative and other diseases outlined above. These animal and cell models may involve over-expression of all or part of PAMP or PAMP mutants, *e.g.*, as mini-gene cDNA transgene constructs under the regulation of suitable promoter elements carried in vectors such as cos-Tet for transgenic mice and pcDNA (Invitrogen, California) in transfected cell lines. Animal and cellular models can also be generated by via homologous recombination mediated targeting of the endogenous gene to create artificially mutant sequences (knock-in gene targeting); or loss of function mutations (knock-out gene targeting); by translocation of P-elements; and by

chemical mutagenesis. Animal, cellular and cell-free model systems can be used for a variety of purposes including the discovery of diagnostics and therapeutics for this disease.

Included within the scope of this invention is a mammal in which two or more genes have been knocked out or knocked in, or both. Such mammals can be generated by repeating the procedures set forth herein for generating each knockout construct, or by breeding to mammals, each with a single gene knocked out, to each other, and screening for those with the double knockout genotype.

Regulated knockout animals can be prepared using various systems, such as the tet-repressor system (*see* US Patent No. 5,654,168) or the Cre-Lox system (*see* US Patents No. 4,959,317 and No. 5,801,030).

Transgenic mammals can be prepared for evaluating the molecular mechanisms of PAMP, and particularly human PAMP/PS1 or PAMP/PS2 function. Such mammals provide excellent models for screening or testing drug candidates. It is possible to evaluate compounds or diseases on “knockout” animals, *e.g.*, to identify a compound that can compensate for a defect in PAMP activity. Alternatively, PAMP (or mutant PAMP), alone or in combination with β APP, PS1, and/or PS2, (double or triple transgenics) “knock-in” mammals can be prepared for evaluating the molecular biology of this system in greater detail than is possible with human subjects. Both technologies permit manipulation of single units of genetic information in their natural position in a cell genome and to examine the results of that manipulation in the background of a terminally differentiated organism. These animals can be evaluated for levels of mRNA or protein expression, processing of β APP, or development of a condition indicative of inappropriate gene expression, *e.g.*, *Notch* phenotype or another phenotype as set forth above, or neurodegeneration, including cognitive deficits, learning or memory deficits, or neuromuscular deficits.

Various transgenic animal systems have been developed. Mice, rats, hamsters, and other rodents are popular, particularly for drug testing, because large numbers of transgenic animals can be bred economically and rapidly. Larger animals, including sheep, goats, pigs, and cows, have been made transgenic as well. Transgenic *D.melanogaster* and *C.elegans* can also be made and, using known genetic methods, may offer the ability to identify upstream and downstream modifiers of a PAMP phenotype. Transgenic animals can also be prepared by introducing the transgene on a vector; such animals, which are not modified in the germ line and are only transiently transgenic, naturally cannot pass along the genetic information to their progeny.

In another series of embodiments, transgenic animals are created in which (i) a human PAMP, or a mutant human PAMP, is stably inserted into the genome of the transgenic animal; and/or (ii) the endogenous PAMP genes are inactivated and replaced with their human

counterparts. See, *e.g.*, Coffman, Semin. Nephrol. 17:404, 1997; Esther *et al.*, Lab. Invest. 74:953, 1996; Murakami *et al.*, Blood Press. Suppl. 2:36, 1996. Such animals can be treated with candidate compounds and monitored for the effects of such drugs on PAMP cavity.

PAMP Gene Therapy

As discussed above, abnormalities in PAMP expression and/or interactions with PS1/PS2/ β APP are associated with severe neurological deficits. Thus, the present invention provides for treatment of such deficits either with a drug discovered using a screening assay or transgenic animal model, or both, as set forth above, or by replacing a defective *PAMP* gene with a functional gene by gene therapy.

A gene encoding PAMP, a PAMP mutant, or alternatively a negative regulator of PAMP such as an antisense nucleic acid, intracellular antibody (intrabody), or dominant negative PAMP (which may be truncated), can be introduced *in vivo*, *ex vivo*, or *in vitro* using a viral or a non-viral vector, *e.g.*, as discussed above. Expression in targeted tissues can be effected by targeting the transgenic vector to specific cells, such as with a viral vector or a receptor ligand, or by using a tissue-specific promoter, or both. Targeted gene delivery is described in International Patent Publication WO 95/28494, published October 1995.

Preferably, for *in vivo* administration, an appropriate immunosuppressive treatment is employed in conjunction with the viral vector, *e.g.*, adenovirus vector, to avoid immuno-deactivation of the viral vector and transfected cells. For example, immunosuppressive cytokines, such as interleukin 12 (IL-12), interferon- γ (IFN γ), or anti-CD4 antibody, can be administered to block humoral or cellular immune responses to the viral vectors (*see, e.g.*, Wilson, Nature Medicine, 1995). In that regard, it is advantageous to employ a viral vector that is engineered to express a minimal number of antigens.

Herpes virus vectors. Because herpes virus is trophic for cells of the nervous system (neural cells), it is an attractive vector for delivery of function *PAMP* genes. Various defective (non-replicating, and thus non-infectious) herpes virus vectors have been described, such as a defective herpes virus 1 (HSV1) vector (Kaplitt *et al.*, Molec. Cell. Neurosci. 2:320-330, 1991; International Patent Publication No. WO 94/21807, published September 29, 1994; International Patent Publication No. WO 92/05263, published April 2, 1994).

Adenovirus vectors. Adenoviruses are eukaryotic DNA viruses that can be modified to efficiently deliver a nucleic acid of the invention to a variety of cell types *in vivo*, and has been used extensively in gene therapy protocols, including for targeting genes to neural cells. Various serotypes of adenovirus exist. Of these serotypes, preference is given to using type 2 or type 5 human adenoviruses (Ad 2 or Ad 5) or adenoviruses of animal origin (*see* WO94/26914). Those adenoviruses of animal origin which can be used within the scope of the present invention include adenoviruses of canine, bovine, murine (example: Mav1, Beard *et*

al., Virology 75 (1990) 81), ovine, porcine, avian, and simian (example: SAV) origin. Preferably, the adenovirus of animal origin is a canine adenovirus, more preferably a CAV2 adenovirus (*e.g.*, Manhattan or A26/61 strain (ATCC VR-800), for example). Various replication defective adenovirus and minimum adenovirus vectors have been described for gene therapy (WO94/26914, WO95/02697, WO94/28938, WO94/28152, WO94/12649, WO95/02697 WO96/22378). The replication defective recombinant adenoviruses according to the invention can be prepared by any technique known to the person skilled in the art (Levrero *et al.*, Gene 101:195 1991; EP 185 573; Graham, EMBO J. 3:2917, 1984; Graham *et al.*, J. Gen. Virol. 36:59 1977). Recombinant adenoviruses are recovered and purified using standard molecular biological techniques, which are well known to one of ordinary skill in the art.

Adeno-associated viruses. The adeno-associated viruses (AAV) are DNA viruses of relatively small size which can integrate, in a stable and site-specific manner, into the genome of the cells which they infect. They are able to infect a wide spectrum of cells without inducing any effects on cellular growth, morphology or differentiation, and they do not appear to be involved in human pathologies. The AAV genome has been cloned, sequenced and characterized. The use of vectors derived from the AAVs for transferring genes *in vitro* and *in vivo* has been described (see WO 91/18088; WO 93/09239; US 4,797,368, US 5,139,941, EP 488 528). The replication defective recombinant AAVs according to the invention can be prepared by co-transfecting a plasmid containing the nucleic acid sequence of interest flanked by two AAV inverted terminal repeat (ITR) regions, and a plasmid carrying the AAV encapsidation genes (*rep* and *cap* genes), into a cell line which is infected with a human helper virus (for example an adenovirus). The AAV recombinants which are produced are then purified by standard techniques.

Retrovirus vectors. In another embodiment the gene can be introduced in a retroviral vector, *e.g.*, as described in Anderson *et al.*, U.S. Patent No. 5,399,346; Mann *et al.*, 1983, Cell 33:153; Temin *et al.*, U.S. Patent No. 4,650,764; Temin *et al.*, U.S. Patent No. 4,980,289; Markowitz *et al.*, 1988, J. Virol. 62:1120; Temin *et al.*, U.S. Patent No. 5,124,263; EP 453242, EP178220; Bernstein *et al.* Genet. Eng. 7 (1985) 235; McCormick, BioTechnology 3 (1985) 689; International Patent Publication No. WO 95/07358, published March 16, 1995, by Dougherty *et al.*; and Kuo *et al.*, 1993, Blood 82:845. The retroviruses are integrating viruses which infect dividing cells. The retrovirus genome includes two LTRs, an encapsidation sequence and three coding regions (*gag*, *pol* and *env*). In recombinant retroviral vectors, the *gag*, *pol* and *env* genes are generally deleted, in whole or in part, and replaced with a heterologous nucleic acid sequence of interest. These vectors can be constructed from different types of retrovirus, such as MoMuLV ("murine Moloney leukemia virus"), MEV ("murine Moloney sarcoma virus"), HaSV ("Harvey sarcoma virus"); SNV

("spleen necrosis virus"); RSV ("Rous sarcoma virus") and Friend virus. Suitable packaging cell lines have been described in the prior art, in particular the cell line PA317 (US 4,861,719); the PsiCRIP cell line (WO 90/02806) and the GP+envAm-12 cell line (WO 89/07150). In addition, the recombinant retroviral vectors can contain modifications within the LTRs for suppressing transcriptional activity as well as extensive encapsidation sequences which may include a part of the gag gene (Bender *et al.*, J. Virol. 61:1639, 1987). Recombinant retroviral vectors are purified by standard techniques known to those having ordinary skill in the art.

Retrovirus vectors can also be introduced by recombinant DNA viruses, which permits one cycle of retroviral replication and amplifies transfection efficiency (*see* WO 95/22617, WO 95/26411, WO 96/39036, WO 97/19182).

Lentivirus vectors. In another embodiment, lentiviral vectors can be used as agents for the direct delivery and sustained expression of a transgene in several tissue types, including brain, retina, muscle, liver and blood. The vectors can efficiently transduce dividing and non-dividing cells in these tissues, and maintain long-term expression of the gene of interest. For a review, *see*, Naldini, Curr. Opin. Biotechnol., 9:457-63, 1998; *see also* Zufferey, *et al.*, J. Virol., 72:9873-80, 1998). Lentiviral packaging cell lines are available and known generally in the art. They facilitate the production of high-titer lentivirus vectors for gene therapy. An example is a tetracycline-inducible VSV-G pseudotyped lentivirus packaging cell line which can generate virus particles at titers greater than 10⁶ IU/ml for at least 3 to 4 days (Kafri, *et al.*, J. Virol., 73: 576-584, 1999). The vector produced by the inducible cell line can be concentrated as needed for efficiently transducing nondividing cells *in vitro* and *in vivo*.

Non-viral vectors. A vector can be introduced *in vivo* in a non-viral vector, *e.g.*, by lipofection, with other transfection facilitating agents (peptides, polymers, etc.), or as naked DNA. Synthetic cationic lipids can be used to prepare liposomes for *in vivo* transfection, with targeting in some instances (Felgner, *et al.*, Proc. Natl. Acad. Sci. U.S.A. 84:7413-7417, 1987; Felgner and Ringold, Science 337:387-388, 1989; *see* Mackey, *et al.*, Proc. Natl. Acad. Sci. U.S.A. 85:8027-8031, 1988; Ulmer *et al.*, Science 259:1745-1748, 1993). Useful lipid compounds and compositions for transfer of nucleic acids are described in International Patent Publications WO95/18863 and WO96/17823, and in U.S. Patent No. 5,459,127. Other molecules are also useful for facilitating transfection of a nucleic acid *in vivo*, such as a cationic oligopeptide (*e.g.*, International Patent Publication WO95/21931), peptides derived from DNA binding proteins (*e.g.*, International Patent Publication WO96/25508), or a cationic polymer (*e.g.*, International Patent Publication WO95/21931). Recently, a relatively low voltage, high efficiency *in vivo* DNA transfer technique, termed

electrotransfer, has been described (Mir *et al.*, C.P. Acad. Sci., 321:893, 1998; WO 99/01157; WO 99/01158; WO 99/01175). DNA vectors for gene therapy can be introduced into the desired host cells by methods known in the art, *e.g.*, electroporation, microinjection, cell fusion, DEAE dextran, calcium phosphate precipitation, use of a gene gun (ballistic transfection), or use of a DNA vector transporter (see, *e.g.*, Wu *et al.*, J. Biol. Chem. 267:963-967, 1992; Wu and Wu, J. Biol. Chem. 263:14621-14624, 1988; Hartmut *et al.*, Canadian Patent Application No. 2,012,311, filed March 15, 1990; Williams *et al.*, Proc. Natl. Acad. Sci. USA 88:2726-2730, 1991). Receptor-mediated DNA delivery approaches can also be used (Curiel *et al.*, Hum. Gene Ther. 3:147-154, 1992; Wu and Wu, J. Biol. Chem. 262:4429-4432, 1987). US Patent Nos. 5,580,859 and 5,589,466 disclose delivery of exogenous DNA sequences, free of transfection facilitating agents, in a mammal.

EXAMPLES

The present invention will be further understood by reference to the following examples, which are provided as exemplary of the invention and not by way of limitation.

EXAMPLE 1: A novel PAMP that mediates β APP processing and *Notch/Glp1* signal transduction

This example shows that both PS1 and PS2 interact with a novel Type I transmembrane protein, PAMP, and that this novel protein also interacts with α - and β -secretase derived fragments of β APP. We also show that abolition of functional expression of the *C. elegans* homologue of the protein leads to a developmental phenotype (anterior pharynx 2 -*aph2*) which is thought to be due to inhibition of the *glp/Notch* signaling pathway. This novel protein is therefore positioned to mediate both the gain of function and loss of function phenotypes associated with presenilin missense mutations and presenilin knockouts, respectively.

Materials and Methods

Antibodies against PS1, PS2 and β APP. An antibody, termed 1142, directed against PS1, was raised to a peptide segment corresponding to residues 30-45 of PS1 (Levesque *et al.*, J Neurochem 1998: 72:999-1008; Yu *et al.*, Biol Chem 1998; 273:16470-16475). The peptide was synthesized by solid-phase techniques and purified by reverse phase high pressure liquid chromatography (HPLC). Peptide antigens were linked to keyhole limpet hemocyanin (KLH) and used, in combination with complete Freud's adjuvant, to inoculate New Zealand White rabbits. Antisera from three rabbits was pooled, ammonium precipitated and the antibody was purified with Sulfo-link (Pierce) agarose-peptide affinity

columns. Other antibodies used include antibody 369, a polyclonal rabbit-anti-human antibody directed against the C-terminus of human β APP (Buxbaum et al., Proc. Natl. Acad. Sci. USA 1990; 87: 6003-6007); antibody 14 (Ab14), a rabbit polyclonal antibody raised against residues 1-25 of human PS1 (Seeger et al., Proc. Natl. Acad. Sci. 1997; 94: 5090-5094); antibody α -PS1-CTF, a polyclonal rabbit antibody directed against the PS1 loop; and antibody DT2, a monoclonal antibody raised to a GST-fusion protein containing the PS2 N-terminal sequence from residues 1-87.

Preparation of presenilin associated components. To identify membrane associated components of the presenilin complex, an immunoaffinity procedure was used to extract PS1 and tightly associated membrane proteins from semi-purified intracellular membrane fractions. Human embryonic kidney cells (HEK) 293 (ATCC) with a stable over-expression of moderate level wild type human PS1, were grown to confluence, washed twice with ice-cold phosphate-buffered saline, and then homogenized with Buffer A (0.25 M sucrose, 20 mM HEPES pH 7.2, 2 mM EGTA, 2 mM EDTA, 1 mM DTT, and a protease inhibitor cocktail containing 5 μ g/ml each of Leupeptin, Antipain, pepstatin A, Chymostatin, E64, Aprotinin, and 60 μ g/ml 4-(2-aminoethyl)-benzenesulfonyl fluoride (AEBSF)). The cell homogenates were centrifuged 1000 \times g for 10 minutes to remove cell debris. The supernatant was then centrifuged 10,000 \times g for 60 minutes. The resulting membrane pellet was resuspended in Buffer B (20 mM HEPES pH 7.2, 1 M KCl, 2 mM EGTA, 2 mM EDTA, 1 mM DTT, and protease inhibitor cocktail as above) and incubated for 45 minutes with agitation at 4°C. Cell membranes were collected again by centrifugation at 107,000 \times g for 60 minutes. The cell membranes were then lysed on ice for 60 minutes with Buffer C (1% Digitonin, 20 mM HEPES pH 7.2, 100 mM KCl, 2 mM EGTA, 2 mM EDTA, 1 mM DTT, and protease inhibitors cocktail). After spinning 10,000 \times g for 15 minutes, the protein extract was adjusted with Buffer C to contain 5 mg/ml protein. A total of 0.5 g of protein was obtained.

Isolation. The extracted proteins were then subjected to fractionation with 10-40% glycerol gradient containing 0.5% Digitonin as described (Yu G, et al., J Biol Chem 1998; 273: 16470-16475). After being verified by Western blotting with anti-PS1 antibodies, the peak fractions containing PS1 were pooled and incubated overnight with Protein A/G agarose coupled with either antibody 1142 or a control IgG purified from preimmune rabbit serum. The Protein A/G agarose beads were washed three times with Buffer D (1% Digitonin, 20 mM HEPES pH 7.2, 100 mM KCl, protease inhibitors cocktail), and three times with Buffer E (0.5% Digitonin, 0.5% CHAPS, 20 mM HEPES pH 7.2, 100 mM KCl, 10 mM CaCl₂, 5 mM MgCl₂, and the protease inhibitor cocktail as above). Isolated protein complexes were eluted from the beads with 0.1M Glycine-HCl, pH 3.0, and then neutralized with 1M Tris. Proteins were then separated by Tris-Glycine SDS-PAGE gels and stained with silver

stain and Coomassie Blue stain. The staining of the immuno-purified proteins displayed two intense bands in addition to those of the presenilin holoprotein and its fragments.

Sequence analysis. Individual protein bands were cut out and analyzed with solid-phase extraction capillary electrophoresis mass spectrometry/mass spectrometry (SPE-CE-MS/MS). Briefly, protein bands were first digested in-gel with trypsin; the digested proteins were extracted and dried in a speed vacuum down to concentrate the peptides; and the peptides thereafter separated with micro LC and analyzed by on-line tandem mass spectrometry (Figeys *et al.*, Anal Chem 1999; 71: 2279-2287). Nucleotide and amino acid sequence homology searches were conducted using the BLAST algorithm, and motif analyses performed using the program BLOCKS.

General transfection and analysis methods. Based on the human PAMP sequence, public databases (*e.g.*, GenBank; World Wide Web (www.ncbi.nlm.nih.gov) were searched for homologous ESTs (SEQ ID NOs: 3-10), which were collected into a few contigs. These contigs all turned out to be correct, but did not cover full-length mouse and *D. melanogaster* cDNAs.

Full length murine (SEQ ID NO: 15), human (SEQ ID NO: 13) and *D. melanogaster* (SEQ ID NO: 17) PAMP cDNAs were obtained using oligonucleotides designed from partial cDNA/EST sequences in public databases to screen appropriate cDNA libraries, for 5'RACE, and/or for RT-PCR experiments. A PAMP expression construct was generated by inserting human PAMP cDNA in-frame with the V5 epitope of pcDNA6 (Invitrogen) at the C-terminus of PAMP. HEK293 cells with a stable expression of PS1/PS2 and β APP_{sw} were transiently transfected with either V5-tagged PAMP or empty plasmid (mock transfection control). Duplicate experiments were performed by: (1) transient transfection of V5-PAMP and β APP₆₉₅ (or empty vector plus β APP₆₉₅ as a mock transfection control) into murine embryonic fibroblasts stably infected with human PS1 expressed from a retroviral vector construct (Clontech, CA); or (2) transient transfection of V5-PAMP (or an empty plasmid) into HEK293 cell lines with a stable expression of the C-terminal 99 amino acids of β APP with a preceding artificial signal peptide (spC100-APP) together with either wild type PS1, PS1-L392V, or PS1-D385A. Cells were lysed with a Digitonin lysis buffer or with 1% NP40, and the protein extracts were subjected to gradient fraction, immunoprecipitation or direct Western blotting as described (Yu G, *et al.*, J Biol Chem 1998; 273: 16470-16475). PS1 was immunodetected or immunoprecipitated with antibodies 14 or α -PS1-CTF; and PS2 was immunodetected or immunoprecipitated with antibody DT2. FL- β APP and its C-terminal α - and β - secretase derivatives were detected using antibody 369.

Results

Isolation of PAMP. Immunoprecipitation of PS1 protein complexes, followed by SDS-PAGE with Coomassie Blue and silver staining, yielded two intense bands in addition to presenilin holoprotein. These bands were characterized by mass spectroscopy analysis.

Mass spectroscopy analysis revealed several *armadillo* repeat containing peptides, (previously known to functionally interact with presenilins (Yu G, *et al.*, J Biol Chem 1998; 273: 16470-16475.; Zhou J, *et al.*, NeuroReport (Fast Track) 1997; 8: 2085-2090; Nishimura M, *et al.*, Nature Med 1999; 5: 164-169), and a novel peptide (PAMP) which had a sequence identified to that predicted for an anonymous, partial cDNA (Genbank; Accession No. D87442). The cDNA sequence predicted an open reading frame of 709 amino acids (SEQ ID NO: 14), which contains a putative N-terminal signal peptide, a long N-terminal hydrophilic domain with sequence motifs for glycosylation, N-myristoylation and phosphorylation, a ~20 residue hydrophobic putative transmembrane domain, and a short hydrophilic C-terminus of 20 residues (Figure 1A and 1B).

Orthologous PAMP proteins. The PAMP amino acid sequence had no significant homology to other proteins within available databases, except for a hypothetical *C.elegans* protein (World Wide Web (www) ncbi.nih.gov; Accession No. Q23316) ($p = 2 \times 10^{-28}$; identity = 22%; similarity = 39%) (SEQ ID NO: 12) ascertained from a genomic DNA sequence (Figure 1A and 1B). In addition to strong primary amino acid sequence conservation, this *C.elegans* protein has a very similar topology to human PAMP, suggesting that it is the nematode orthologue of human PAMP.

In the absence of functional clues arising from homologies to other known proteins, the predicted amino acid sequences of the murine (SEQ ID NO: 16) and *D.melanogaster* (SEQ ID NO: 18) orthologues of PAMP were cloned and examined. The four orthologous PAMP proteins had a similar topology and significant sequence conservation near residues 306-360, 419-458, and 625-662 of human PAMP (SEQ ID NO: 14) (Figure 1A and 1B). Motif analysis of these conserved domains revealed a weak similarity (strength = 1046) between residues 625 – 641 (ARLARALSPAFELSQWS; SEQ ID NO: 19) of mouse and human PAMP to cyclic nucleotide binding domains. While the putative transmembrane domain sequences were not highly conserved, all four orthologues contained a conserved serine residue within this hydrophobic domain. Finally, there were four conserved cysteine residues in the – terminal hydrophilic domain (Cys₁₉₅, Cys₂₁₃, Cys₂₃₀, and Cys₂₄₈ in human PAMP) which had a periodicity of 16-17 residues in the N-terminus, and may form a functional domain (*e.g.*, a metal binding domain or disulfide bridges for stabilizing the tertiary structure of PAMP/PAMP complexes).

Interaction of PAMP with presenilin 1. To confirm the authenticity of the PAMP:PS1 interaction, HEK293 cells were transiently transfected with PAMP cDNA (SEQ ID NO: 13) tagged at the 3'-end with a V5-epitope encoded from the pcDNA6 vector. The conditioned media were collected 20 hr after transient transfection with PAMP (or with empty vector), and the A β ₄₀ and A β ₄₂ levels were measured by ELISA (Zhang L, *et al.*, J Biol Chem 1999; 274: 8966-8972.). In Western blots of lysates of these cells, the use of anti-V5 (Invitrogen, CA) and enhanced chemiluminescence (Amersham) detected a V5-immunoreactive band of ~110 kDa which was reduced to ~80 kDa following Endo H digestion (equivalent to the size predicted from the PAMP amino acid sequence), confirming the predicted glycosylation of PAMP. In addition, a series of about 7-10 kDa fragments were observed, which are predicted to contain the TM domain and short C-terminus of PAMP plus the 3 kDa V5- epitope. These C-terminal derivatives may be authentic cleavage products of full-length PAMP, or, alternatively, a proteolytic artifact arising from the attachment of the V5-epitope to the C-terminus of PAMP.

Reciprocal immunoprecipitation studies in cells expressing combinations of transfected V5-tagged-PAMP, transfected wild type or mutant PS1, transfected wild type PS2, or endogenous presenilins, confirmed the PS1:PAMP interaction, and showed a similar interaction between PAMP and PS2. In contrast, immunoprecipitation of other ER-resident proteins (*e.g.*, calnexin) failed to show any evidence of an interaction between these proteins and PAMP. Glycerol velocity gradient analysis of the native conformation of PAMP revealed that PAMP was co-eluted into the same high molecular weight fractions as PS1 and PS2, indicating that it is an authentic component of the high molecular weight presenilin protein complexes. These biochemical data were supported by immunocytochemical studies, which showed that transfected PAMP and endogenous PS1 strongly co-localized in the ER and Golgi in MDCK canine kidney/epithelial cells (ATCC). Similar studies with PS2 confirmed that PAMP also tightly associates with both endogenous PS2 in human brain and with transfected PS2 in HEK293 cells.

The PAMP gene. Chromosomal locations and genetic map positions of the murine and human PAMPS were obtained from public genetic and transcriptional maps (World Wide Web ([www](http://www.ncbi.nlm.nih.gov)) [ncbi.nlm.nih.gov](http://www.ncbi.nlm.nih.gov)). The gene encoding PAMP is located on human chromosome 1 near the genetic markers D1S1595 and D1S2844. The 5'- end of the PAMP gene is embedded in the 5'- end of the coatmer gene encoded on the opposite strand. The human PAMP gene is close to a cluster of markers which have yielded positive, but sub-significant evidence for linkage to or association with Alzheimer Disease in two independent genome wide surveys (Kehoe P, *et al.* Hum Mol Genet 1999; 8: 237-245). The murine PAMP maps within a 700 Kb interval of murine chromosome 1 which contains the gene defect

associated with *Looptail* phenotype in mice (Underhill DA, *et al.*, Genomics 1999; 55: 185-193). Mice heterozygous for *Looptail* show developmental defects in dorsal axial structures including notochord, brain, spinal cord, and somites (Greene ND, *et al.*, Mech Dev 1998; 73: 59-72.), which are reminiscent of those observed in PS1^{-/-} mice (Shen J, *et al.*, Cell 1997; 89: 629-639; Wong PC, *et al.*, Nature 1997; 387:288-292). These observations suggest that the presenilin: PAMP complex may be involved in both β APP and *Notch* processing.

***C. elegans* homolog of PAMP.** The *C. elegans* homolog of PAMP corresponds to the *aph-2* gene. Mutations in *aph-2* have been identified in a screen for mutants with phenotypes identical to embryonic mutant phenotypes caused by loss of *glp-1* activity, *i.e.*, lack of an anterior pharynx, e.g. cDNA clone. The EST corresponding to *aph-2*, (cDNA clone yk477b8, kindly provided by Y. Kohara, National Institute of Genetics, Japan) was sequenced and the coding region (SEQ ID NO: 11) found to match exactly the Genefinder prediction made by the *C. elegans* sequencing consortium (Genbank; Accession No. Z75714). Double stranded RNA interference (RNAi) confirmed the mutant phenotype of *aph-2*. Sense and antisense RNA were transcribed *in vitro* from PCR product amplified from the phage yk477b8. After annealing equal quantities of sense and antisense products, the dsRNA product made was injected into L4 stage wild-type worms. The chosen line of worms, designated lin-12(n302) (Greenwald and Seydoux, Nature 1990: 346:197-199; Greenwald, *et al.*, Cell 1983: 34; 435-444) was obtained from the Caenorhabditis Genetics Center. Injected animals were transferred to fresh plates daily and the progeny scored at least 36 hours after injection for the embryonic lethal phenotype and 4-5 days after injection for the egg-laying phenotypes. Animals injected with dsRNA from yk477b8 template produced eggs that lacked an anterior pharynx. These results support the notion that *aph-2*/PAMP contributes to cell interactions mediated by *glp-1*/*Notch* in the embryo.

Functional role for the PAMP:presenilin complexes in β APP processing. To examine a functional role for the PAMP:presenilin complexes in β APP processing, the interactions between PAMP, PS1, and β APP, and its derivatives were investigated. The cell lines used were transiently transfected with V5-tagged PAMP, and stably expressing wild type β APP₆₉₅ in addition to wild type PS1, wild type PS2, PS1-L392V mutant, or PS1-D385A mutant. The PS1-L392V mutation is a pathogenic mutation associated with familial AD (Sherrington R, *et al.*, Nature 1995; 375:754-760) and with increased secretion of A β ₄₂ (Scheuner D, *et al.* Nature Med 1996; 2: 864-870, Citron M, *et al.* Nature Med 1997; 3: 67-72). The PS1-D385A mutation is a loss of function mutation associated with inhibition of PS1 endoproteolysis and a decrease in γ -secretase activity (Wolfe MS, *et al.*, Nature 1999; 398: 513-517). The conditioned media were collected 20 hr after transient transfection with PAMP (or with empty vector), and the A β ₄₀ and A β ₄₂ levels were measured by ELISA (Zhang

L, *et al.*, J Biol Chem 1999; 274: 8966-8972). Analysis of Western blots from these co-immunoprecipitation experiments revealed that PAMP holoprotein (and C-terminally tagged proteolytic fragments of PAMP) interacted in equivalent degrees with wild type PS1, wild type PS2, PS1-L392V mutant, and PS1-D385A mutant proteins. In addition, PAMP holoprotein and the C-terminal proteolytic fragments of PAMP also co-immunoprecipitated with the C-terminal proteolytic fragments of β APP but not β APP holoprotein in lysates of cells expressing either β APP holoprotein or just the C-terminal 99 amino acids of β APP. Significantly, compared to cells expressing equivalent quantities of wild type PS1, cell lines expressing pathogenic mutations of PS1 showed increased amounts of C-terminal β APP fragments co-immunoprecipitating with PAMP. Conversely, cell lines expressing the loss-of-function PS1-D385A mutation showed greatly reduced amounts of C-terminal β APP derivatives co-immunoprecipitating with PAMP despite the presence of very large amounts of C-terminal β APP derivatives in these cells.

These results were confirmed in HEK293 cells over-expressing either β APP^{Swedish} or the SpC99- β APP cDNA. The latter encodes the C-terminal 99 residues of β APP (corresponding to the products of γ -secretase cleavage) plus the β APP signal peptide. The interaction of PAMP appears much stronger with C99- β APP than that with C83- β APP. However, C83- β APP is much less abundant in these cells (Fig. 6b, middle panel, lanes 1-4). In fact, PAMP does interact with both C99- and C83- β APP stubs (see Fig. 6c, lane 9 and Fig. 8d). Cumulatively, these results indicate that PAMP likely interacts with the C-terminal derivatives of β APP which are the immediate precursors of A β and p3. However, of greater interest, the genotype of the co-expressed PS1 molecule dynamically influenced the interaction between PAMP and C99-/C83- β APP stubs. Thus, more C-terminal β APP fragments co-immunoprecipitated with PAMP in cells expressing the FAD-associated PS1-L392V mutation compared to cells expressing wild type PS1 (and equivalent quantities of nicastrin and C99- β APP). Conversely, much less C-terminal β APP derivatives co-immunoprecipitated with PAMP in cell lines expressing the loss-of-function PS1-D385A mutation (despite the presence of very large amounts of C-terminal β APP derivatives in these cells). These effects are more easily seen in cells over-expressing the C99- β APP construct. However, similar but less pronounced differences were also observed in cells over-expressing full-length β APP^{Swedish}. More importantly, the PS1-sequence-related differences in the interaction of PAMP with C-terminal β APP derivatives were most evident within 24 hours of transient transfection of PAMP. By 72 hours, the PS1-sequence-related differences were largely abolished. This dynamic change in the interaction of PAMP with C99/C83- β APP was not due to changes in the levels of PS1, C-terminal β APP derivatives or PAMP. One interpretation of these results is

that the presenilins may be dynamically involved in regulating or loading PAMP with the substrates of γ -secretase.

Presenilin binding domains of PAMP. In transiently transfected cells (in which the 7-10 kDa C-terminal of PAMP can be detected), anti-PS1 immunoprecipitation products contain both full length PAMP and the \sim 7-10 kDa C-terminal PAMP fragments. Similarly, in these cells, immunoprecipitation with antibodies to the C-terminus of β APP (Ab369) also renders C-terminal nicastrin epitopes. The TM domain of PAMP is not highly conserved in evolution. These results suggest that the C99-/C83- β APP and PS1/PS2-binding domain(s) of PAMP are in the TM domain or C-terminus.

Discussion

The above results indicate that PAMP is a component of the PS1 and PS2 intracellular complexes. The observations that PAMP also binds to the C-terminal fragment of β APP (arising from α - and β -secretase cleavage of full length β APP), that the degree of binding of these fragments to PAMP is modulated by mutations in PS1, and that the direction of this modulation is congruent with the effects of each mutant of A β production (*i.e.*, the pathogenic L392V mutation increases binding to PAMP and increases A β ₄₂ production whereas the D385A mutation has the opposite effects) strongly argues that PAMP is part of a functional complex involved in processing of C-terminal β APP derivatives. Similarly, the observation that inhibition of PAMP expression in *C. elegans* leads to a phenotype similar to that of *glp/Notch* loss of function, argues that PAMP, like PS1 and PS2, is also a functional component of the pathways involved in processing of *Notch*. This conclusion is strengthened by the fact that the murine PAMP gene maps within a 700 kb interval on murine chromosome 1 which carries the *Looptail* mutant, and is thus likely to be the site of the *Looptail* mutation. *Looptail* has a number of phenotypic similarities to those of *Notch* and PS1 knockouts in mice. Because *Looptail* is a model of human spinal cord malformations including spina bifida, PAMP biology may also provide some useful insights into this neurological developmental defect as well.

At the current time the exact role of PAMP in the presenilin-complex-mediated processing of β APP and *Notch*-like molecules is not fully defined. Inspection of the primary amino acid sequence of PAMP does not reveal very strong homologies to known proteases. However, the recombinant expression systems of the invention permit evaluation of three-dimensional structure of PAMP; it is possible that PAMP itself has a protease activity. However, it is currently more plausible that PAMP plays another role in β APP and *Notch* processing. Thus, PAMP may be involved in the function of PS1 and PS2 complexes by binding substrates for γ -secretase. The efficacy of this binding is clearly modulated by PS1

EXAMPLE 2: PAMP mutants useful for studies on Alzheimer's Disease

Site-directed mutagenesis was used to generate the following artificial mutations in PAMP:

Cys: PAMP_{C230A} in the 4 conserved cystine motif
DYIGS: PAMP_{D336A/Y337A} in the central conserved region
D369L: PAMP_{Δ312-369} in the central conserved region
D340X: PAMP_{Δ312-340} in the central conserved region
YDT: PAMP_{D458A} in the putative 'aspartyl protease' DTA site
SPAF: PAMP_{P633A/F635A} in the SPAF motif
TM: PAMP_{S683A} in the TM domain
C3D: PAMP_{Δ630-668} in the conserved region adjacent to the TM domain

To further examine the role of nicastrin in β APP processing, we inserted PAMP cDNAs, harboring the above mutations as well as normal/wild type PAMP (PAMP_{wt}) cDNA and the cDNA for an unrelated protein (LacZ), in frame into pcDNA6 vectors. A series of HEK293 cell lines stably expressing endogenous PS1, β APP_{Swedish} and either wild type nicastrin or nicastrin constructs in which various conserved domains had been mutated or deleted, were then created by transfection. PAMP expressing cells were selected with lacticidin to generate stable cell lines. Conditioned media from these cell lines were collected after 6-24 hours and $A\beta_{40}$ and $A\beta_{42}$ were measured by ELISA.

In the PAMP_{D336A/Y337A} mutant, both $A\beta_{40}$ and $A\beta_{42}$ levels were increased, and there was also a 68% increase in $A\beta_{42}/A\beta_{40}$ ratio which is very similar to that observed in clinical mutations in β APP, PS1, and PS2, associated with early onset Alzheimer Disease. The $A\beta_{42}/A\beta_{40}$ ratio was also increased in one cell line expressing the PAMP_{C230A} mutant.

In contrast, both the total $A\beta_{42}$ and $A\beta_{40}$ levels and the $A\beta_{42}/A\beta_{40}$ ratio were massively reduced (to only 18% of the control) in the PAMP_{Δ312-369} mutant. A similar but less profound reduction of both the total $A\beta_{42}$ and $A\beta_{40}$ levels and the $A\beta_{42}/A\beta_{40}$ ratio was observed in the conditioned medium from the PAMP_{Δ312-340} cell lines.

There is no apparent difference in $A\beta_{42}$ or $A\beta_{40}$ levels, or in the $A\beta_{42}/A\beta_{40}$ ratio, when the PAMP_{wt}, PAMP_{D458A}, PAMP_{Δ630-668}, PAMP_{P633A/F635A}, and PAMP_{S683A} cells were compared to control lines (expressing LacZ, or empty vector).

Thus, certain PAMP mutants cause biochemical changes similar to those induced by mutations in the β APP, PS1, and PS2 genes which give rise to Alzheimer Disease. These artificial PAMP mutations can therefore be used to generate cellular and other model systems to design treatments and preventions for Alzheimer Disease related disorders. These

mutations also show that PAMP is involved in the pathogenesis of AD, and may provide information for the design of specific molecular diagnostics or therapeutics.

When compared to mock-transfected or LacZ transfected cells, overexpression of wild type PAMP, and overexpression of most PAMP mutation- or deletion-constructs had no significant effect on A β secretion. However, missense mutation of the conserved DYIGS motif to AAIGS (residues 336-340 of human PAMP) caused a significant increase in A β ₄₂ secretion, a smaller increase in A β ₄₀ secretion, and an increase in the A β ₄₂/A β ₄₀ ratio ($p < 0.001$; Table 2). This increase in A β ₄₂ production was equivalent to that of FAD-related missense mutations in PS1. Conversely, deletion of the DYIGS domain in two independent constructs (PAMP _{Δ 312-369} and PAMP _{Δ 312-340}) caused a significant reduction in both A β ₄₂ and A β ₄₀ secretion which was more profound in PAMP _{Δ 312-369} cells than in PAMP _{Δ 312-340} cells (Table 2). The magnitude of the reduction in A β secretion in PAMP _{Δ 312-369} cells was equivalent to that observed with the PS1-D385A loss-of-function mutation. Somewhat unexpectedly, and in contrast to PS1^{-/-} and PS1-D385A cells, the reduction in A β secretion in NCT _{Δ 312-369} and NCT _{Δ 312-340} cells was not accompanied by the expected accumulation of C99- and C83- β APP stubs. Since there was no consistent change in the levels of soluble β APP (β APP_s) in the conditioned medium of any of the PAMP mutant cells, the most probable explanation for this result is that C99- and C83- β APP stubs which do not enter the PAMP:presenilin complex for γ -secretase cleavage to A β may be degraded by other pathways.

The effects of PAMP mutations on A β secretion were not due to trivial explanations such as differences in the levels of PAMP, β APP holoprotein, or PS1/PS2. None of these mutations caused any consistent, detectable change in the amount of APP_s in conditioned medium or in the amount of C99/C83- β APP that could be co-immunoprecipitated with PAMP. However, both the PAMP _{Δ 312-369} mutant and the PAMP _{Δ 312-340} deletion mutant significantly reduced the amount of PS1 which could be co-immunoprecipitated with PAMP. Interestingly, the reduction in efficiency of binding to PS1 was proportional to the reduction in A β secretion induced by each deletion mutant. Multiple mechanisms underlying the effect of mutations in the first conserved domain can explain these results. This domain contains no obvious functional motifs (*e.g.*, for glycosylation etc.), nor does it have significant sequence homology to other known proteins. Consequently, the three functionally active PAMP mutations either affect a presenilin-binding domain in PAMP, or affect a specific regulatory domain of PAMP which modulates both direct binding of PAMP to PS1 and the subsequent γ -secretase-mediated cleavage of PAMP-bound C99- and C83- β APP stubs.

Table 2

Transfection	Normalized A β ₄₂	Normalized A β ₄₀	A β ₄₂ /A β ₄₀ Ratio
Mock (LacZ/empty vector)	1.0	1.0	1.0
wild type nicastrin	1.03 \pm 0.09	1.05 \pm 0.07	0.99 \pm 0.07
D336A/Y337A	3.09 \pm 0.59 (p < 0.001)	1.61 \pm 0.19 (p = 0.001)	1.81 \pm 0.15 (p < 0.001)
nicastrin ₃₁₂₋₃₆₉	0.05 \pm 0.04 (p < 0.001)	0.31 \pm 0.06 (p < 0.001)	0.09 \pm 0.05 (p < 0.001)
nicastrin ₃₁₂₋₃₄₀	0.33 \pm 0.04 (p = 0.002)	0.55 \pm 0.04 (p = 0.001)	0.59 \pm 0.06 (p = 0.003)

The present invention is not to be limited in scope by the specific embodiments described herein. Indeed, various modifications of the invention in addition to those described herein will become apparent to those skilled in the art from the foregoing description and the accompanying figures. Such modifications are intended to fall within the scope of the appended claims.

It is further to be understood that values are approximate, and are provided for description.

Patents, patent applications, and publications are cited throughout this application, the disclosures of which are incorporated herein by reference in their entireties.